Application Note

085/2012





Digest Automat K-438 KjelMaster K-375 with KjelSampler K-376 Nitrogen and Protein Determination in Animal Feed According to the Kjeldahl Method



Nitrogen and Protein Determination in Animal Feed According to the Kjeldahl Method

The determination of protein in feed is a routine procedure for quality assurance and labelling. A simple and fast method for protein determination in feed products according to the ISO 5983-2:2009 and AOAC 954.01 regulations is introduced below. The sample is digested with sulfuric acid and Kjeldahl Tablets Titanium using the digest Automat K-438, followed by distillation and titration with the KjelMaster K-375 with KjelSampler K-376.

Introduction

Protein determination is one of the key analyses performed in the feed industry. The samples require digestion with sulfuric acid to convert nitrogen into ammonium sulfate. After conversion to ammonia through the alkalinization with sodium hydroxide, the sample is distilled into a boric acid receiver by steam distillation, followed by a titration with sulfuric acid solution. The nitrogen content is multiplied by a sample-specific protein factor (6.25 for feed products) to obtain the protein content.

Experimental

Instrumentation:

Digest Automat K-438 KjelMaster K-375 with KjelSampler K-376

Samples:

Dog food with a labelled protein content of 11 g/100 g, rabbit food with a labelled protein content of 15 g/100 g and cat food with 37 g/100 g.

Determination:

The samples were added directly into a sample tube as described in Table 1. A portion of 15 ml sulfuric acid, two titanium tablets and one antifoam tablet were added, and the digestion was performed using the parameters specified in Table 2. After digestion the ammonia of the sample was distilled into a boric acid solution by steam distillation and titrated with sulfuric acid (Table 3).

The method was verified by using 0.15 g tryptophane as a reference substance.

Table 1: Weight for each sample

| | Sample weight [g] |
|-------------|-------------------|
| Dog food | 1.0 |
| Rabbit food | 1.0 |
| Cat food | 0.5 |

Table 2: Temperature profile for digestion with the K-438

| Step | Temp. [°C] | Time [min] |
|---------|------------|------------|
| Preheat | 320 | - |
| 1 | 420 | 120 |
| Cooling | - | 30 |

Table 3: Parameters for distillation and titration using the KjelMaster K-375 with KjelSampler K-376

| with rejeled inplet it of | | | | | |
|---------------------------|---------------|--------------------|---|--|--|
| Method Parameters | | | | | |
| H ₂ O volume | 50 ml | Steam output | 100% | | |
| NaOH volume | 60 ml | Receiving solution | 30 ml H₃BO₃ 4% | | |
| Reaction time | 5 s | Titration solution | H ₂ SO ₄ 0.1 mol/l | | |
| Dist. mode | Fixed time | Endpoint pH | 4.65 | | |
| Dist. time | 180 s | Stirrer sp. titr. | 7 | | |
| Stirrer sp. dist. | 5 | Titr. algorithm | Optimal | | |

Results

The tryptophane recoveries were 100.0 %, rsd = 0.2 % (n=4). The determined protein contents are presented in Table 4.

| Table 4: Determined | protein contents i | (rsd in | brackets | n=4) | |
|---------------------|--------------------|----------|-----------|-------|--|
| | protein contents i | (130 111 | Diackets, | 11-4) | |

| Product | Labelled protein content | Determined protein content |
|-------------|--------------------------|-------------------------------|
| Dog food | 11 g/100 g | 10.45 g/100 g (0.2%) |
| Rabbit food | 15 g/100 g | 14.79 g/100 g (0.5%) |
| Cat food | 37 g/100 g | 38.40 g/100 g (0.2%) |

Conclusion

The determination of protein contents in feed products using the K-438 and KjelMaster K-375 with KjelSampler K-376 provides reliable and reproducible results that correspond to the labelled values with low relative standard deviations. The total digestion time is approx. 120 min.

References

ISO 5983-2:2009 AOAC 954.01

Operation manual DigestAutomat K-432/ K-438 Operation manual KjelMaster K-375 with KjelSampler K-376

For more detailed information and safety considerations please refer to Application Note 085/2012.

1 Introduction

An easy and reliable method for determining nitrogen and protein contents in feed products according to the Kjeldahl method, as described in the ISO 5983-2:2009, and AOAC 954.01 regulations, is introduced below. The samples are digested using the Digest Automat K-438. The distillation and boric acid titration are performed using the KjelMaster K-375 with KjelSampler K-376.

2 Equipment

- Digest Automat K-438 (the parameters used are also valid for K-437)
- Scrubber K-415 TripleScrub ^{ECO}
- KjelMaster K-375 with KjelSampler K-376
- Mixer, Retsch Grindomix GM200
- Analytical balance (accuracy ± 0.1 mg)

3 Chemicals and Materials

- Sulfuric acid conc 98 %, Fluka (84727)
- Titanium, BUCHI Kjeldahl Tablet (11057980)
- Antifoam, BUCHI Kjeldahl Tablet (11057984)
- Sodium hydroxide 32 %, Brenntag (81980-452)
- Boric acid 4 %, 200 g of boric acid, Brenntag (80948-155) diluted to 5 l with deionized water, pH adjusted to 4.65
- Sulfuric acid 0.1 mol/l, Fluka (35357) standard solution
- Neutralization solution for the Scrubber: 600 g of sodium carbonate, calcined, technical, Synopharm (0179420) about 2 ml of ethanol and a spatula tip of bromthymol blue, Fluka (18460) diluted to 3 l with distilled water
- DL-Tryptophan (Assay >99 %), Alfa Aesar (L05936)
- Sucrose, Riedel-de Haën (16104)

For a safe handling please pay attention to all corresponding MSDS data sheets!

Samples:

- Dog food with a labelled protein content of 11 g/100 g
- Rabbit food with a labelled protein content of 15 g/100 g
- Cat food with a labelled protein content of 37 g/100 g

The samples were purchased at a local supermarket.

The samples were mixed for 10 s up to visual homogeneity.

4 **Procedure**

The determination of nitrogen and protein contents in feed products includes the following steps:

- Homogenization of the samples
- Digestion of the samples, with the Digest Automat K-438
- Distillation and titration of the samples, using the KjelMaster K-375 with KjelSampler K-376

4.1 Digestion method - tryptophane (verification of the method)

- Preheat the Digest Automat K-438 according to the parameters listed in Table 2
- Place approx. 0.15 g of tryptophane and 0.7 g sucrose into a 300 ml sample tube
- Add two Titanium Tablets, one Antifoam Tablet and 15 ml sulfuric acid (98 %)
- Prepare additional blanks, 2 ml water and 0.7 g of sucrose as a sample
- Carefully suspend the sample by gently swirling the tube
- Connect the Scrubber K-415 to the Digest Automat K-438 for absorbing the acid fumes created during the digestion
- Insert the rack containing the samples into the lift and immediately start the digestion according to the parameters listed in Table 2

4.2 Digestion method - samples

- Preheat the Digest Automat K-438 according to the parameters listed in Table 2
- Place each sample into a 300 ml sample tube as described in Table 1

| Sample | Weight [g] |
|-------------|------------|
| Dog food | 1.0 |
| Rabbit food | 1.0 |
| Cat food | 0.5 |

Table 1: Weight for each sample

- Add 2 Titanium tablets, one Antifoam Tablet and 15 ml of sulfuric acid (98 %) to each tube
- Prepare additional blanks, 2 ml water and 0.7 g of sucrose as a sample
- Connect the Scrubber K-415 to the K-438 for absorbing the acid fumes created during digestion
- Insert the rack containing the samples into the lift and immediately start the digestion according to the parameters listed in Table 2

| Table 2. Temperature profile for digestion with the K-438 | | | | |
|---|---------------------|-----|--|--|
| Step | ep Temperature [°C] | | | |
| Preheating | 320 | - | | |
| 1 | 420 | 120 | | |
| Cooling | - | 30 | | |

Table 2: Temperature profile for digestion with the K-438

- If the liquid inside the sample tube is not clear and blue-green, digest for an additional 30 min at 420 °C.
- Let the samples cool down

NOTE: The samples should be clear-green immediately after the digestion. A darkening of the clear liquid samples during the cooling down process is possible but does not affect the results.

4.3 Distillation and titration

Distill the samples according to the parameters listed in Table 3

| Method parameters KjelMaster K-375 | | | | |
|------------------------------------|---|-------------------------|----------------|--|
| H ₂ O volume | 50 ml Titration solution H ₂ SO ₄ 0.1 mol/l | | | |
| NaOH volume | 60 ml | Sensor type | Potentiometric | |
| Reaction time | 5 s | Titration mode | Standard | |
| Distillation mode | Fixed time | Measuring mode | Endpoint pH | |
| Distillation time | 180 s | Endpoint pH | 4.65 | |
| Stirrer speed distillation | 5 | Stirrer speed titration | 7 | |
| Steam output | 100 % | Titration start volume | 0 ml | |
| Titration type | Boric acid | Titration algorithm | Optimal | |
| Receiving solution vol. | 30 ml | | | |

Table 3: Distillation and titration using the KjelMaster K-375 with KjelSampler K-376

4.4 Calculation

The results are calculated as a percentage of nitrogen. In order to calculate the protein content of the sample, the nitrogen content is multiplied with a sample-specific protein factor. The following equations (1), (2), and (3) are used to calculate the results.

$$w_{N} = \frac{(V_{Sample} - V_{Blank}) \cdot z \cdot c \cdot f \cdot M_{N}}{m_{Sample} \cdot 1000}$$
(1)

$$%N = w_N \cdot 100 \%$$
 (2)

$$\% P = w_N \cdot PF \cdot 100 \%$$
 (3)

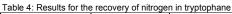
| : weight fraction of nitrogen |
|--|
| : amount of titrant for the sample [ml] |
| : mean amount of titrant for the blank [ml] |
| : molar valence factor (1 for HCl, 2 for H_2SO_4) |
| : titrant concentration [mol/l] |
| : titrant factor (for commercial solutions normally 1.000) |
| : molecular weight of nitrogen (14.007 g/mol) |
| : sample weight [g] (recovery: consider the assay of tryptophan) |
| : conversion factor [ml/l] |
| : percentage of weight of nitrogen |
| : percentage of weight of protein |
| : sample-specific protein factor (6.25 for feed products) |
| |

5 Results

5.1 Recovery of tryptophane

The results of the nitrogen determination and recovery in tryptophane are presented in Table 4. The nominal value of tryptophane (assay: \geq 99.0 %) is 13.72 % nitrogen (the value corrected with the assay is 13.58 % nitrogen). The recoveries are within the specification of > 98.5 % [1].

| Tryptophan | m _{Sample} [g] | V _{Sample} [ml] | %N | Recovery [%] |
|------------|-------------------------|--------------------------|--------|--------------|
| Sample 1 | 0.1508 | 7.504 | 13.578 | 99.9 |
| Sample 2 | 0.1538 | 7.662 | 13.601 | 100.1 |
| Sample 3 | 0.1536 | 7.618 | 13.538 | 99.7 |
| Sample 4 | 0.1576 | 7.840 | 13.589 | 100.0 |
| Average | - | - | 13.577 | 100.0 |
| Rsd [%] | - | - | 0.2 | 0.2 |



The mean blank volume for this sample was 0.195 ml (n = 4).

5.2 Protein determination in feed products

The results of the determination of nitrogen in feed products are presented in Tables 5 - 7.

| Table 5: Results for the determination of a | nitrogen in dog food | I (declared protein con | tent 11 g/100 g) |
|---|----------------------|-------------------------|------------------|
| | | | |

| Dog food | m _{Sample} [g] | V _{Sample} [ml] | %N | %P |
|----------|-------------------------|--------------------------|-------|-------|
| Sample 1 | 1.0672 | 6.580 | 1.676 | 10.48 |
| Sample 2 | 1.0460 | 6.438 | 1.672 | 10.45 |
| Sample 3 | 1.0405 | 6.388 | 1.667 | 10.42 |
| Sample 4 | 0.9751 | 6.024 | 1.675 | 10.47 |
| Average | - | - | 1.672 | 10.45 |
| Rsd [%] | - | • | 0.2 | 0.2 |

The mean blank volume for this sample was 0.195 ml (n = 4).

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|---|---|--|
| Table 6: Results for the determination of nitrogen in rabbit for | od (declared protein content 15 d/100 d) | |
| Table 6. Recalls for the determination of mategori in fabble los | ou (ucoluicu protein content to griec g) | |

| Rabbit food | m _{Sample} [g] | V _{Sample} [ml] | %N | %P |
|-------------|-------------------------|--------------------------|-------|-------|
| Sample 1 | 1.0240 | 8.772 | 2.346 | 14.67 |
| Sample 2 | 1.0374 | 8.994 | 2.376 | 14.85 |
| Sample 3 | 0.9979 | 8.620 | 2.365 | 14.78 |
| Sample 4 | 1.0072 | 8.744 | 2.378 | 14.86 |
| Average | - | - | 2.366 | 14.79 |
| Rsd [%] | - | - | 0.5 | 0.5 |

The mean blank volume for this sample was 0.195 ml (n = 4).

| Cat food | m _{Sample} [g] | V _{Sample} [ml] | %N | %P |
|-----------|-------------------------|--------------------------|-------|-------|
| Sample 1 | 0.5105 | 11.365 | 6.129 | 38.31 |
| Sample 2 | 0.4940 | 11.030 | 6.144 | 38.40 |
| Sample 3 | 0.5212 | 11.660 | 6.162 | 38.51 |
| Sample 4 | 0.5005 | 11.164 | 6.139 | 38.37 |
| Average - | | - | 6.144 | 38.40 |
| Rsd [%] | - | - | 0.2 | 0.2 |

Table 7: Results for the determination of nitrogen in cat food (declared protein content 37 g/100 g)

The mean blank volume for this sample was 0.195 ml (n = 4).

6 Comparison to Standard Methods

This application note is based on the standard methods ISO 5983-2:2009 and AOAC 954.01. The standard method AOAC 954.01 does only use 2 g sucrose as blanks.

Table 8: Differentiation from the standard methods

| | This application note | Standard methods | Notes/Impact |
|---------------------|--|---|---|
| Catalyst | 2 titanium tablets (each 3.5 g K_2SO_4 ; 0.11 g CuSO ₄ ; 0.11 g TiO ₂) | ISO: 2 tablets (each 3.5 g K ₂ SO ₄ ; 0.4 g CuSO ₄) AOAC: 15 g K ₂ SO ₄ ; 0.65 g Hg | Easy to handle especially in routine analytics. The choice of catalyst does not influence the result. No toxic Hg. |
| Sulfuric acid | 15 ml | ISO: 12 - 15 ml AOAC: 25 ml | No impact. Same ratio of sulfuric acid / catalyst |
| Water | 50 ml | ISO: 80 ml AOAC: 200 ml | The K-375 generates steam in a separate vessel; therefore, it is not necessary to add such a high amount of water to the digested sample as described in the standard methods. |
| Sodium hydroxide | 60 ml (Conc.: 32%) | ISO: 50 ml (Conc.: 40 %) AOAC: 80 ml (Conc.: 32 %) | No impact. Same ratio of sodium hydroxide / sulfuric acid. Sodium hydroxide 32% is more gentle to the pump than higher concentrated alkali. The sample solution must be strongly alkaline. |
| Receiving solution | 30 ml boric acid solution 4 % | ISO: 25 - 30 ml boric acid solution 4 % AOAC: 0.5 N H ₂ SO ₄ (Back titration) | No impact. The boric acid titration and the back titration lead to equal results. |
| Titration solution | H ₂ SO ₄ 0.2 N | ISO: HCI 0.1 N AOAC: 0.1 N NaOH (Back titration) | No impact. The volume of titration solution should be between 3 and 17 ml. |

7 Conclusion

The determination of nitrogen and protein in feed products using the Digest Automat K-438, KjelMaster K-375, and KjelSampler K-376 provides reliable and reproducible results that correspond to the labelled values with low relative standard deviations. The recoveries with tryptophane were 100.0 % and are within the specification of > 98.5 % [1].

The most important characteristics of the latest version of the fully-automatic Kjeldahl sampler system, KjelMaster K-375 with KjelSampler K-376, are the convenient and intuitive operation, short process time as well as sophisticated data management. In combination with the Digest Automat K-438, personnel attendance during sample analysis is significantly reduced.

8 References

[1] ISO 5983-2:2009 AOAC 954.01

Operation manual DigestAutomat K-432/ K-438 Operation manual of Scrubber B-415 Operation manual KjelMaster K-375 with KjelSampler K-376

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